

16

11/16/94

Project No. _____
Book No. _____

TITLE Deep Vent / GAPDH / dT primers

From Page No. _____

Purpose: Since GAPDH - PCR worked with 3' Thiol primers attempted the same amplification with other available primers, under same conditions.

Deep Vent buffer
200 µg Terytaké
200 µM dNTP
1 µM primers

enzymat 1 U and 0.5 U
Mg ab. 2, 3, 4 and 6 mM

did just one of each:

* 2697 & 2696 no dU Lac FWD & Lac Reverse (100 µM)
 " " " 3' PPT
 + dU " " (100 µM)

each primer set - rxn Rx were made.

Regular	17 - 20 = 0.5 U	dU	23 - 36 = 0.5 U	3' - 1 PPT
	21 - 24 = 1.0 U		37 - 40 = 1.0 U	25

10x buffer	50	50	50	50	29
dNTP	10	10	10	10	
primer 1	5	5	5	5	20
2	5	5	5	5	20
Template	20 (100 µg/2)	20	20	20	20
H ₂ O	860	270	270	270	330

150 → 45 µl/Rx ← 450

added 5µl of	2	3	4	6 mM	57.200
Mg chelation	1	1	1	1	
ml	0	0.5	1	2 (100 mM)	10 x
	5	4.5	4	3 11.0	

enzymat added individually
0.25 µl for 0.5 U
0.5 µl for 1.0 U

To Page No. _____

Witnessed & Understood by me,

Date

11/20/94

Inv. nt d by

R cord d by

Dr. Schaeffer

Dat

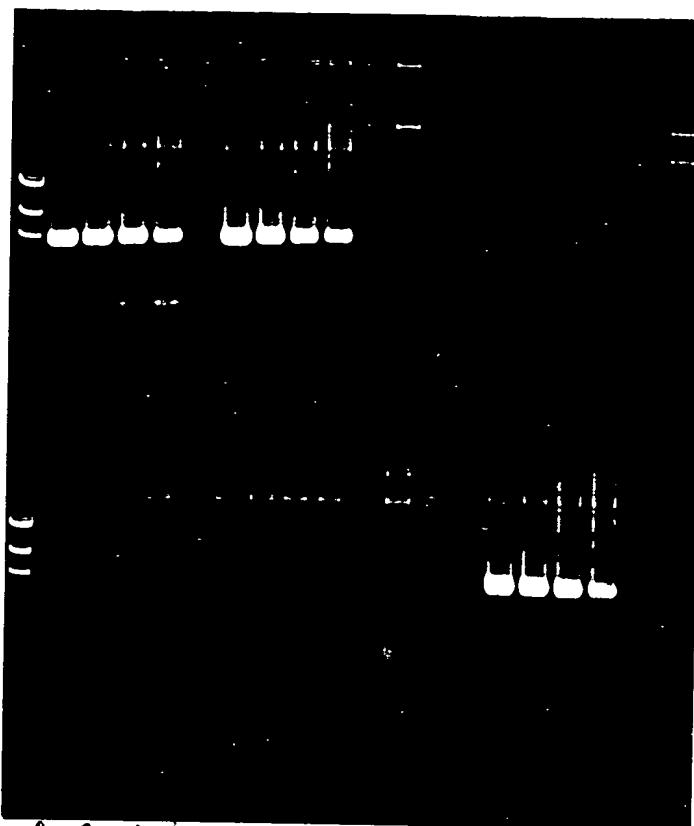
11/16/94

ag N

Regular

3'-1 PPT

0.5U



Result:

Regular - unmodified
modified - so could not
say 3'-1 PPT is better
than unmodified!

dv certainly has problems
with Deep Vent.

With the assumption of bands
seen with 1' I don't know
why there is no banding
with 0.5U & 3'-1 PPT
primers.

Can I template - ?
Circumstances -
what are these bands on
the top.

Template / primer - no
enzyme controls also
have these?

Vent 7/ - exo didn't discriminate
between modified and
unmodified
with dv in earlier this year a 500 bp (ac. 2) never got
amplified

- samples dried 12/19/94

To Page No. _____

ed & Understood by m .

Date

Invented by

Dat

11/18/94

Recorded by

R. S. Sitaarman

11/18/94